

Fractional Order Impedance Model to Estimate Glucose Concentration: in Vitro Analysis

Dana Copot, Robin De Keyser, Jasper Juchem, Clara M. Ionescu

Ghent University, Research group on Dynamical Systems and Control

Technologiepark 914, 9052 Ghent, Belgium

{dana.copot,claramihaela.ionescu,robain.dekeyser,jasper.juchem}@ugent.be

Abstract: This paper uses tools from fractional calculus such as Cole-Cole and fractional order impedance models for estimation of glucose concentration. The measured impedance is compared with two fractional order models and the simulation results show that Cole-Cole model has limitation and cannot capture the dynamics of the simulated environment. On the other hand, the fractional order model can follow the changes in impedance for several study cases. Model parameters are correlated with various conditions of the test environment. The results of these study cases show that the fractional order model is a suitable candidate for this particular application. The hypothesis tested in this paper provides new tools for glucose concentration monitoring and measurement.

Keywords: Fractional Order Impedance Models; Diabetes; Modeling; Cole-Cole Model

1 Introduction

Due to the increase in diabetes severity, this disease has become a serious health problem at a worldwide level representing the main cause of death in the age category 20-79 years. Hence, the research community has focused on developing more reliable and accurate devices for glucose measurement [1]. In this research area the main challenge to tackle is the design of non-invasive devices. Although, the non-invasive idea dates already from the 70s many of the proposed technologies are still at an early stage of development [2].

Several techniques such as electromagnetic sensing, infrared spectroscopy, polarimetry were considered in order to enable the design of new methodologies for glucose concentration measurement. Worldwide, this area is intensively explored and efforts are being made towards the development of stable, reliable and accurate measurement devices (e.g. BioSensors, Cnoga Medical C8 MediSensors, etc.).

Although there are several devices available on the market, they are not yet working at the desired capacity and there is ongoing research for upgrade. Moreover, the documentation released by the research groups is limited and also validation of this devices on large groups is still missing [3]. The main disadvantage of the current

devices is that they are relatively poor in signal-to-noise-ratio. Therefore, up to now no device can be considered fully reliable and clinically validated. Given that non-invasive techniques are still at an early stage of development there is still room for improvements [3].

Therefore, focus on developing systems such as artificial pancreas to control insulin delivered to patients with diabetes has increased. The current standards in clinical practice consist of blood glucose measurements based on finger-prick technique. However, when this method is used for most patients is an unpleasant procedure due to the protocol of taking blood samples. Moreover, recent studies show that on long term there is the risk of fingerprint damage [4]. In figure 1 an overview of the methods used for blood glucose monitoring and control are presented.

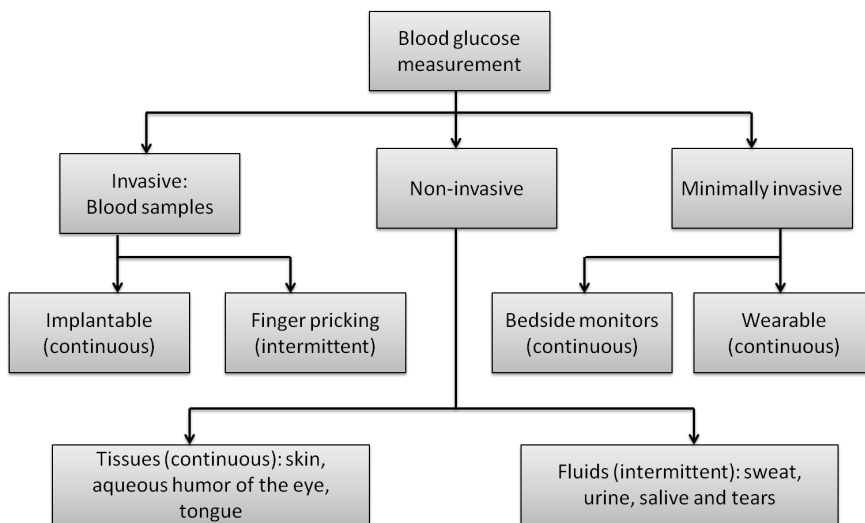


Figure 1

An overview of the method used for blood glucose measurement

Considering all the aspects presented above the advantages of non-invasive technology is easily motivated. Nowadays, the use of nano-technology is the new approach towards the improvement of the health sector. For example, nanorobots are used to deliver a drug to a precise location. This will result in a faster effect and also will reduce the risks of secondary effects. Blood can be characterized as a non-newtonian fluid and therefore when designing such a device this aspect needs to be taken into account [5]. In order to deal with difficulties encountered (from a modeling and control approach) new techniques need to be investigated.

Therefore, this paper discusses the use of fractional calculus since it is able to make a trade off between linear theory (Newtonian) and non-linear theory (Non-Newtonian) fluids.

More specifically, fractional order impedance is a suitable tool to estimate, measure and evaluate the glucose levels. FC is of key importance in modeling of phenomena in several areas of research (e.g. chemistry, physics, engineering, etc.) [6, 7]. From

the perspective of biomedical engineering fractional order calculus has been only recently introduced [8].

In the biomedical engineering research community the interest on using tools from fractional calculus in this area has increased considerably in the last years [9–11]. Fractional order concept refers to dynamical systems for which the structure consist of arbitrary-order derivatives and integrals [12]. A detailed description of fractional order integrals and derivatives can be found in [13].

In this paper the hypothesis wheter or not fractional calculus is a suitable candidate to measure, estimate and evaluate the level of glucose. For this, electrochemical impedance spectroscopy has been employed and preliminary analysis (in vitro) has been performed. EIS is used in several fields of research (e.g. batteries, fuel cells, biological systems, etc.) [14–18] EIS has been succesfully applied to estimate and analyze the impact of glucose in erythrocytes [19, 20], effect of glucose in aqueous solutions [21, 22], etc. This paper proposes a new approach, i.e. the use of fractional order impedance models to characterize changes in glucose concentration (in vitro analysis). At the same time, analysis on how the model parameters are changing as a function of the concentration will be also investigated. The preliminary results indicate that the proposed fractional order model can detect and differentiate changes in glucose concentration.

The paper is organized as follows: In section II A brief description of the tools proposed in this paper is given followed by Section III where the measurement device and the methodology employed are introduced. The outcome of the proposed hypothesis is given in section IV. The main outcome of this study is summarized in section V.

2 Theoretical Background

2.1 Cole-Cole Impedance Model

Fractional order elements are present in many areas of engineering [23]. When an electrochemical system is modelled usually a constat phase element is chosen since a non-homogeneous surface is present [23]. Impedance (when a CPE is used) is obtained using the following formula:

$$Z(j\omega) = \frac{1}{C_\alpha(j\omega)^\alpha} \quad (1)$$

where C_α represents the capacitance of order α , with $0 \leq \alpha \leq 1$.

Electrical properties of cell membrane have been modelled by means of impedance measurements in the 1940 by Cole. Recently, the work of Magin presented a generalization of the Cole-Cole expression by means of fractional calculus [16, 17]. This model consist of 3 main elements: a low frequency resistor R_0 , a high frequency resistor R_∞ and a CPE_α [24].

The developed circuit can be visualised as two serial connected elements: one is R_∞ and the second one is $(R_0 - R_\infty) || CPE_\alpha$. The complex impedance described by Cole

model is given by the following equation (Cole 1940):

$$Z_\alpha(\omega) = R_\infty + \frac{R_0 - R_\infty}{1 + (j\omega\tau_\alpha)^\alpha} \quad (2)$$

with τ_α the characteristic relaxation time.

Previous work of the authors have also employed Cole-Cole model for characterization of several simulated environments (i.e. simulating different concentrations of glucose) and the obtained results are presented in [24].

2.2 Fractional Order Impedance Model

The Cole-Cole model has several limitations (as described in [24]) in terms of capturing the dynamics of the real data. Next, tools from fractional calculus have been employed and an augmented fractional order model has been used to characterize the glucose concentration profile. Given the structure of the fractional order model, electrical analogy with ladder networks can be considered [10]. It has been shown that the values of the complex impedance are increasing as a function of the number of neural networks.

A conceptual scheme of such a ladder network is depicted in the following Figure. The total admittance of such a ladder is given by the continued fraction expansion.

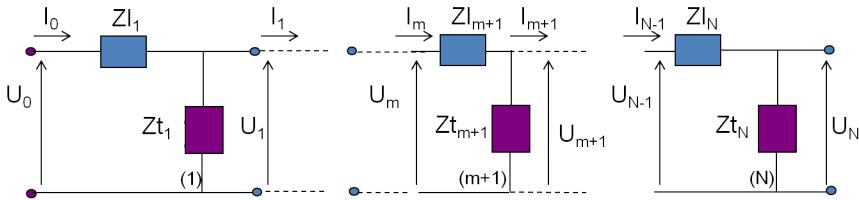


Figure 2

Schematic representation of a ladder network characterizing the neuron transmission model [10]

$$Y_N(s) = \frac{1/Zl_1}{1 + \frac{Zt_1(s)/Zl_1(s)}{1 + \frac{Zt_1(s)/Zl_2(s)}{1 + \frac{Zt_2(s)/Zl_3(s)}{\ddots}}}} \quad (3)$$

$$1 + \frac{Zt_{N-1}(s)/Zl_N(s)}{1 + Zt_N(s)/Zl_N(s)}$$

In previous work of the authors it has been demonstrated that this structure of the ladder network results in the appearance in lumped form of a fractional order parameter dependent on the structure and functionality of the ladder elements [10]:

$$\gamma = \frac{\log(a)}{\log(a+b)} \quad (4)$$

where a and b represent the specific property parameters. Then, the lumped admittance model can be expressed as:

$$Y(s) \approx Ks^\gamma \quad (5)$$

or equivalently, the impedance:

$$Z(s) \approx \frac{k}{s^\gamma} \quad (6)$$

When interpreting the model at a micro scale level, the model refers to the diffusion process of glucose molecules in the simulated environment. Independently, the lumped impedance model has been shown to be of the form:

$$Z(s) = Ds^\delta \quad (7)$$

where D represents the diffusion parameter and δ represents the fractional order parameter value related to the diffusion rate. The complete fractional order model is expressed as:

$$Z(j\omega) = R + \frac{k}{s^\gamma} + Ds^\delta \quad (8)$$

When this model is employed there are 5 parameters which need to be estimated for every impedance measurement.

The reasoning behind developing such a model is to enable the design and development of a nanoscale robot for continuous evaluation of glucose levels. Such a technology would lead to a better monitoring of patients with diabetes by means of an accurate and reliable tool.

3 Experimental Setup and Methodology

3.1 Solutions

To test the proposed methodology, simulated solutions of different glucose concentration have been used for analysis. It is well known that the composition of plasma consists in 92% water and dissolved solutes of 8% such as electrolytes, proteins, organic wastes and nutrients. Electrolytes are responsible with the transmission of electrical signals. The major electrolytes in plasma are Sodium (Na^+), Potassium (K^+) and Chloride (Cl^-) and these are responsible to ensure stable concentrations and to charge differences across cell membranes. For the study cases presented in this paper aqueous solution of KCl with a concentration of 80 mmol/L have been used. In order to simulate the real life situation this baseline solution has been diluted in other 5 concentration levels, i.e. 5, 10, 15, 20 and 40 mmol/L. To ensure that the analysis performed is as close as possible to real life the sample concentration have been chosen based on the data presented in Table 1.

Table 1
Overall glucose level and the medical interpretation.

mmol/L	mg/dL	Medical interpretation
2.0	35	very low, danger of unconsciousness
3.0	55	marginal insulin reaction
4.0 - 6.0	70 -100	normal value before meal in nondiabetic patient
8.0	150	normal value after meal in nondiabetic patient
10.0	180	maximum after meal in nondiabetic patient
15	270	high to very value
16.5 - 20.0	300 - 360	danger
22	400	extremely high

In clinical practice the following glucose levels have been defined (recommended): prior a meal (preprandial) it is indicated that the level of glucose is less than 100 mg/dL (5.5 mmol/L) in plasma and 89 mg/dL (4.9 mmol/L) in whole blood capillary; after meal (postprandial) the value cannot exceed 140 mg/dL (7.8 mmol/L) in plasma and 125 mg/dL (6.9 mmol/L) in whole blood or capillary as shown in table 1, based on which the samples concentration has been selected.

3.2 Instrumentation

Impedance measurements have been performed using an electrochemical device i.e. a Solatron modulabXm impedance analyzer (Solatron Analytical, UK), see Figure 3. The experimental procedure consist in sending a sinusoidal voltage to the electrodes connected to the analyzed sample. The impedance has been evaluated in the frequency interval 1Hz-1MHz. The measurement cell is a small cylindrical tank (inner diameter of 20 mm and a length of 30 mm) and the electrodes used to aquired the measured data are screen-printed electrodes. These electrodes are suitable for repeatable measurements but also they exhibit high electrochemical activity.

3.3 Methodology

Mostly, the electrochemical impedance spectroscopy method is done by means of a single frequency. However, this a time-consuming procedure and it is more difficult to eliminate the transient. Another possibility is to use a random signal which will decrease the amount of time required for analysis, but, in this situation leakage might introduce error in the frequency response function estimation [21]. Therefore, it is important to preserve linearity and to ensure this small amplitude signals need to be send to the analyzed probe and this will result in a poor signal to noise ration. To overcome these multisine excitation signals are a suitable choice. These signal are able to ensure a short measurement time due to their ability to perform broadband excitation. Also, in case of multisine signals leakage is not an issue since it is a periodic signal. For the analysis performed in this paper a multisine excitation signal

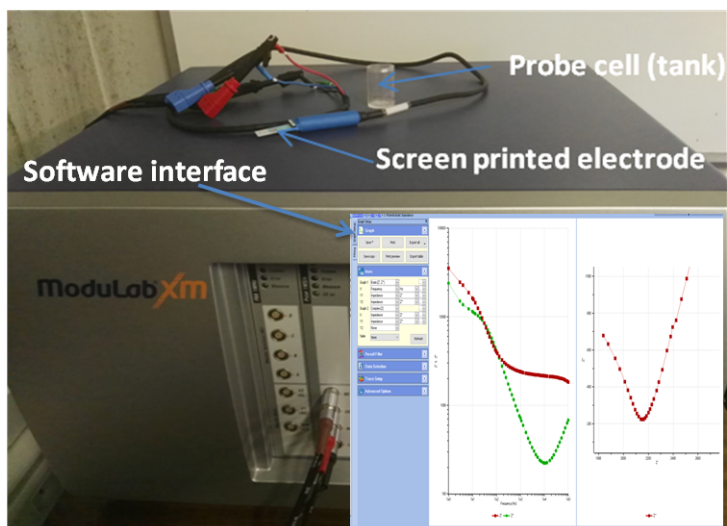


Figure 3
Solatron ModulabXm impedance analyzer

(i.e. voltage) has been sent to the probe and the corresponding current signals have been acquired. This method has the advantage of proving a good analysis of the noise and at the same time of the bias present in the analyzed data. This enables extraction of the best linear approximation from the measured data. For identification, the non-parametric best linear approximation method has been employed. More details about this method can be found in [10]. The outcome of the identification procedure is a complex impedance which can be expressed as real and imaginary part for each excited frequency point. This is a graphical interpretation of the analyzed data which allows us to extract a parametric representation. More specifically, the extracted data is compared with the parametric model described in section 2. This results in a parameterization of the analyzed data. In previous work the authors showed that the method employed in this paper is equally good as a full optimization procedure [10]. In Figure 3 a typical measurement output is depicted.

4 Results

In this section the experimental results obtained for different glucose concentration solutions are presented and discussed. Using the device described in section 2 and employing the methodology described in the paper analysis of multiple simulated solutions has been performed and the results are presented in figures 4-10. The reasoning of the analysis performed in this paper has been described in 3.2. First, the measured data has been fitted to the Cole-Cole model. For this, it is necessary to identify the values of the five model parameters (R_0 , R_∞ , τ , α , β) for the Cole-Cole model. This model proved not to be able to capture the dynamics of measured impedances. In

figure 4 the results obtained for concentration of 5mmol/L and 10mmol/L glucose are presented.

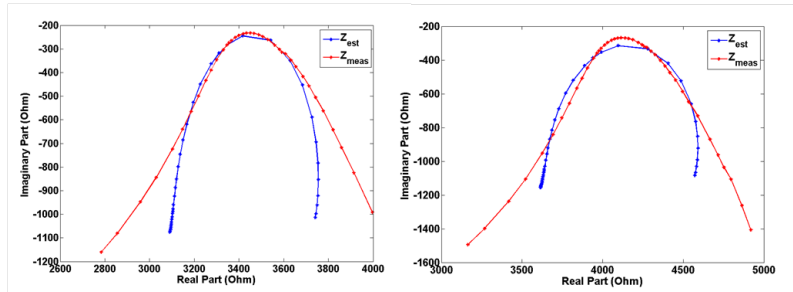


Figure 4

Results of the measured and estimated impedance using Cole-Cole approach. Blue line: model; Red line: measured data. Left: 0 mmol/L glucose; Right: 5 mmol/L. The measurements were performed in the frequency range 1Hz-0.1MHz

As it can be noticed from Figure 4 the obtained results are not satisfactory. More specifically, it can be seen that the typical ellipsoid shape of the estimated impedance does not comply with the somewhat skewed polar plot of the calculated impedance from voltage-current measurements. Therefore, we have moved one step further and tools from fractional calculus have been employed.

For the fractional order impedance model the following parameters have to be estimated R , L , D , α , β . The identified values for every parameter are given in table 2. From table 2 it can be noticed that three model parameters are well characterized while two parameters are yet to be identified. For these two parameters the right experiment has not been yet found but there is ongoing work. The real and imaginary part of the measured impedance for the in vitro tests performed are shown in Figure 5.

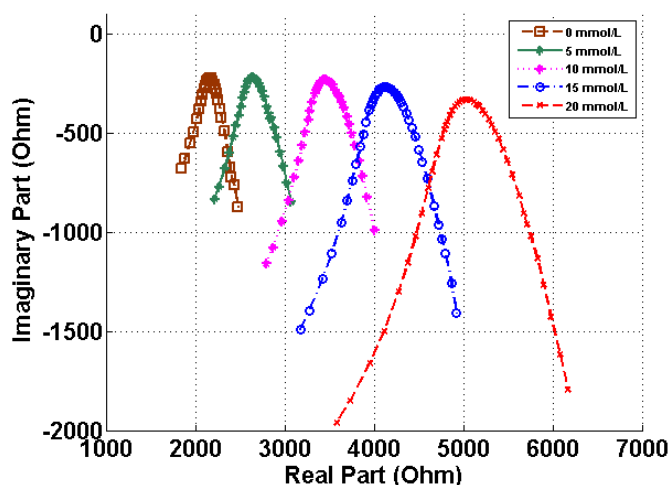


Figure 5

Relevant part of the measured signal used for further analysis

Table 2
Fractional order impedance model parameters estimation.

R	L	D	α	β
0.1512	0.2862	2.9583	0.0017	0.00009
0.1568	0.1700	2.6854	0.00011	0.00009
0.1658	0.0180	2.1128	0.0013	0.00009
0.2164	0.1210	2.6358	0.00012	0.00009
0.2358	0.3521	2.2657	0.0015	0.00009
0.2688	0.0130	1.8598	0.00015	0.00009

In Figures 6-10 the results obtained for different glucose concentration are presented and it can be noticed that a good fit between measured and estimated impedance has been obtained. As it can be noticed from the model parameters also, there is still room for improvement and research with respect to the model performance is ongoing. More specifically, tests to understand the changes of the impedance at high frequencies as well as why at higher concentration a good fitting is not obtained anymore are being investigated.

Conclusions

In this paper the concept of fractional calculus and electrochemical impedance spectroscopy has been used to estimate and measure the impedance of several aqueous glucose solutions. The results show that the electrochemical impedance spectroscopy can be used as a basis for detection of glucose. However, there is still room for improvement in order to obtain a better correlation between the estimated and measured impedance.

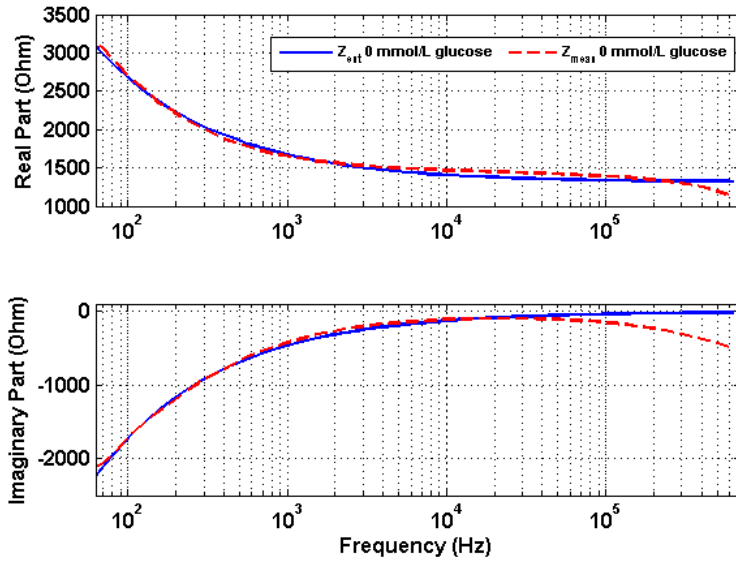


Figure 6

Model (o) against measured impedance (+) at a concentration of 0 mmol/L glucose in the frequency range 1Hz-0.1 MHz

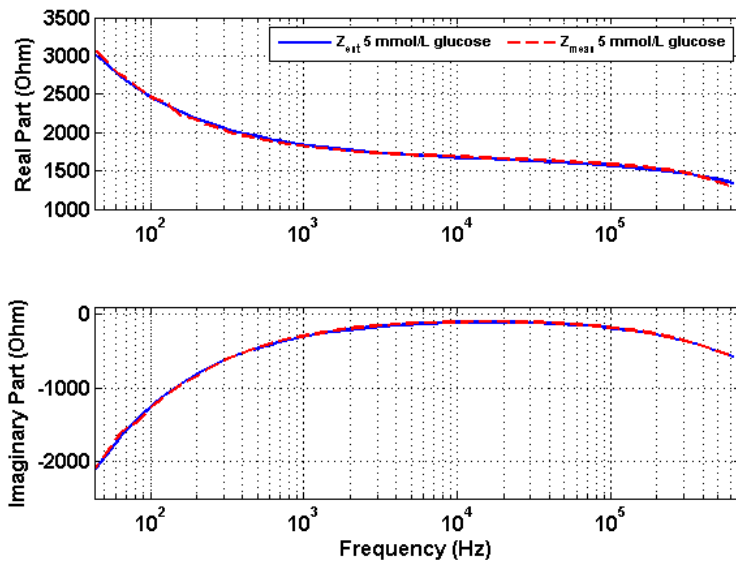


Figure 7

Model (o) against measured impedance (+) at a concentration of 5 mmol/L glucose in the frequency range 1Hz-0.1 MHz

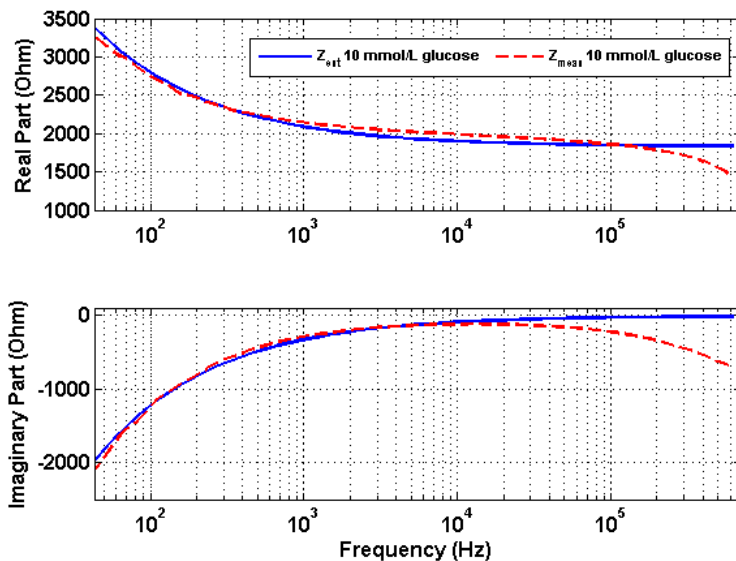


Figure 8

Model (o) against measured impedance (+) at a concentration of 10 mmol/L glucose in the frequency range 1Hz-0.1 MHz

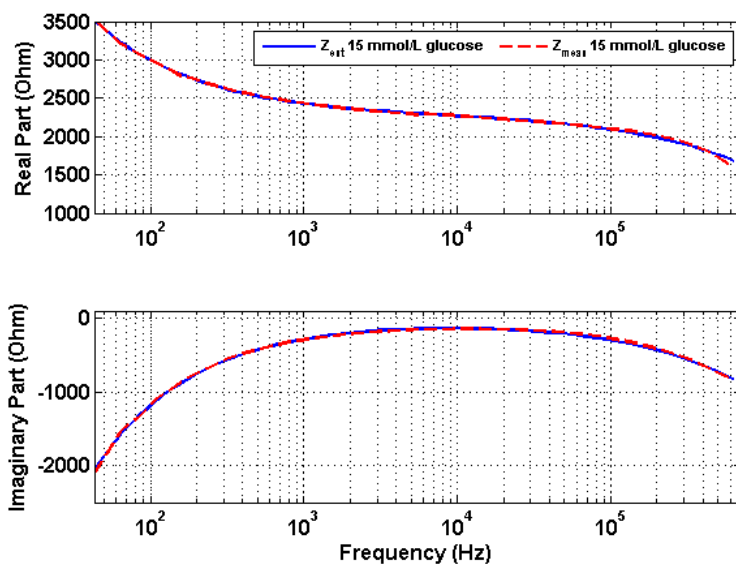


Figure 9

Model (o) against measured impedance (+) at a concentration of 15 mmol/L glucose in the frequency range 1Hz-0.1 MHz

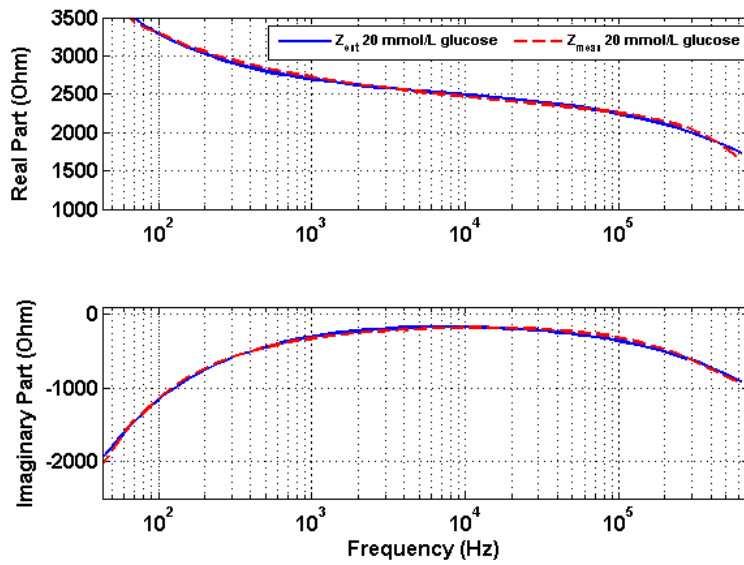


Figure 10

Model (o) against measured impedance (+) at a concentration of 20 mmol/L glucose in the frequency range 1Hz-0.1 MHz

Acknowledgement

This work is financially supported by Flanders Research Centre, grant nr 12B3415N, G008113N and G026514N.

References

- [1] S. Vashist, Continuous glucose monitoring systems: A review. *Diagnostics*, 3, 385-412, 2013.
- [2] O. Khalil, Non-invasive glucose measurement technologies: an update from 1999 to the dawn of the new millennium. *Diabetes Technology Therapy*, 6(5), 660–697, 2004.
- [3] C. So, K. Choi, T. Wong, J. Chung, Recent advances in noninvasive glucose monitoring. National Center for Biotechnology Information, doi:10.2147/MDER.S28134, 2012.
- [4] V. Sgurev, R. Yager, J. Kacprzyk and K. Atanassov. Recent contributions in intelligent systems. Springer International Publishing Switzerland, 2017.
- [5] C. Ionescu, A memory-based model for blood viscosity. *Communications in Nonlinear Science and Numerical Simulation*, 45, 29-34, 2017.
- [6] R. L. Magin. *Fractional Calculus in Bioengineering*, Begell House Publishers, 2006.
- [7] E. Gutierrez, J. Mauricio Rosario and J. T. Machado. *Fractional Order Calculus: Basic Concepts and Engineering Applications*, Mathematical Problems in Engineering, 2010.
- [8] B. J. West. *Fractal physiology and chaos in medicine*, Studies of nonlinear phenomena in life sciences, vol.1. Singapore: World Scientific, 1990.
- [9] G. A. Losa, D. Merlini, T. F. Nonnenmacher and E. R. Weibel. *Fractals in Biology and Medicine*, vol IV. Berlin: Birkhauser, 2005.
- [10] C. Ionescu, *The human respiratory system: an analysis of the interplay between anatomy, structure, breathing and fractal dynamics*, Springer, Series in BioEngineering ISBN 978-1-4471-5387-0, Online ISBN 978-1-4471-5388-7, 2013.
- [11] C. Ionescu, Phase Constancy in a Ladder Model of Neural Dynamics. *IEEE TRANSACTIONS ON SYSTEMS MAN AND CYBERNETICS PART A-SYSTEMS AND HUMANS*, 42(6), 1543-1551, 2012.
- [12] A. Oustaloup. *La derivation non entiere*. Hermes, Paris, 1995. (in French)
- [13] I. Podlubny. *Fractional differential equations*, San Diego: Academic Press, 1999.
- [14] F. Mansfeld, W.J. Lorenz, Electrochemical impedance spectroscopy; applications in corrosion science and technology, in R. Varma, J.R. Selman (Eds.) , *Techniques for Characterization of Electrodes and Electrochemical Processes*, Wiley, New York, 581-647, 1999.
- [15] A. Lasia, Electrochemical impedance spectroscopy and its applications, in R.E. White, B.E. Conway, J.O'M. Bockris (Eds.), *Modern Aspects of Elec-*

- trochemistry, vol. 32, Kluwer Academic/Plenum Press, New York, 143-248, 1999.
- [16] R. Magin, Fractional calculus in bioengineering 1. Critical Reviews in Biomedical Engineering, 32, 1-104, 2004.
- [17] R. Magin, Fractional calculus in bioengineering 2. Critical Reviews in Biomedical Engineering, 32, 105-194, 2004
- [18] H. Fricke, Theory of electrolytic polarization. Philosophical Magazine, ed. Taylor and Francis, 14, 310-318, 1932.
- [19] R. Lisin, B. Ginzburg, M. Schlesinger, Y. Feldman, Time domain dielectric spectroscopy study of human cells. I. Erythrocytes and ghosts. Biochimica et Biophysica Acta, 1280(1), 34-40, 1996.
- [20] A. Caduff, L. Livshits, Y. Hayashi, Y. Feldman, Cell Membrane Response on d-Glucose Studied by Dielectric Spectroscopy. Erythrocyte and Ghost Suspensions. Journal of Physical Chemistry, 108(6), 13827-13830, 2004.
- [21] Park, J.H. Kim, C.S. Choi, B.C. Ham, Biosens. Bioelectron. 19 (4), 321–324
- [22] A. Tura, S. Sbrignadello, S. Barison, S. Conti, G. Pacini, Impedance spectroscopy of solutions at physiological. Journal of Physical Chemistry glucose concentrations, 129(2-3), 235-241, 2007.
- [23] E. Barsoukov, J. Macdonald, Impedance spectroscopy theory, Experiment and Applications. New Jersey: John Wiley and Sons, 595p, 2005.
- [24] D. Copot, C. Muresan. R. De Keyser, C. Ionescu, Fractional order modeling of diffusion processes: a new approach for glucose concentration estimation. International Conference on Automation, Quality and Testing, Robotics, Cluj Napoca, Romania, 19-21 May, 2016 (accepted).
- [25] R. Kadara, N. Jenkinson, C. Banks, Characterisation of commercially available electrochemical sensing platforms. Sensors and Actuators B: Chemical, 138(2), 556-562, 2009.